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Contribution to nanomaterials safety assessment: the need for integrating *in vitro, in vivo* and *in silico* strategies

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The number of NANOMATERIALS brought to market has exponentially grown in recent years and will continue to grow and evolve to new generation NMs.

NANOTECHNOLOGIES – Key-enabling technologies that use materials and manipulations at the nanoscale and which has a potential influence on almost any technological area.

Fundamental and application-driven research is expected to boost nanosciences and innovation towards development of SAFE-BY-DESIGN NMs and applications

NANOMATERIALS (NMs) and NANOTECHNOLOGIES (III)

Nano-applications in consumer products, medicine and industrial processes are widespread – **SOCIETAL BENEFITS**



WHO, 2012

Human exposure to NMs is growing very fast but

Solid information about hazard is limited for the majority of NMs, especially related to chronic exposure to low doses, that is the most likely to occur (e.g., through consumers products)



Zhao & Liu, 2012

The genotoxic effects of NMs, which may be linked to carcinogenic effects, are of special concern because cancer has a long latency period and thereby these effects can be less obvious and more difficult to predict than eventual acute effects.

NANO-BIO INTERACTIONS AND POTENTIAL EFFECTS



Assessment of toxicity - use of complementary *in silico, in vitro* and *in vivo* assays, taking into account specific physico-chemical properties of NMs

Factors interfering with genotoxicitity tests interpretation and comparability

Incomplete description of the NMs physicochemical properties

Coating

Dynamic behavior of NMs (formation of aggregates and agglomerates, and the kinetics dependent of the medium conditions)

Corona formation and composition

Dosing

(difficult to picture a real exposure scenario in *in vitro* or *in vivo* assays - limited human exposure data)

- Interference with colorimetric assays
 - (e.g., cytotoxicity assays)
- Differences in the means of dispersion of insoluble NMs
- Different uptake capacity of cell lines
- Limited existence/access of SOPs and validated methods

The dose-metrics

(e.g., mass, particle number or surface area)

Lack of reliable positive controls at the nanoscale

IN VITRO AND IN VIVO GENOTOXICITY ASSESSMENT OF TIO, NMS

• Pigment

•Food colorant •Cosmetics

> •Skin care products •Sunscreen products

•Photocatalytic properties

- Solar panels
- paints and construction products



Objectives

To assess genotoxic effects of TiO₂ NMs at cellular, molecular and organism level using a combination of *in vitro* and *in vivo* approaches to allow an integrated understanding of its biological effects

Solution Minimize variability inherent to NMs and *in vitro* experimental procedures:

- Benchmark NMs (JRC repository)
- Characterized physico-chemical properties
- Standardized method for NMs dispersion and control of particle size distibution
- MN assay (OECD guideline 487)
- Comet assay (SOP)

Use of integrated *in vivo* approach:

analysis of several endpoints in the same animals (3Rs)
DNA and chromosome damage and somatic gene mutation; inflammation and NPs accumulation in liver (toxicokinetics information)

Comparison of *in vitro* and *in vivo* data for one TiO2 NM



Experimental strategy

Titanium dioxide nanomaterials (JRC repository)



NM-103

NM-104

NM-105

Characterization of physico-chemical properties – **TEM**¹

Nanomaterial		Phase (and other	Impurities/	Specific surface area	Primary particles				Aggregates/agglomerates ^d		
		information)*	coatings (surface modification)	(m²/g) *	Feret Min ± SD (nm) ^b	Feret Max ± SD (nm) ^b	Aspect ratio ± SD ^b	N ^c	25% (nm)	Median (nm)	75% (nm)
TiO ₂	NM- 102	Anatase	-	90	20.8 ± 1.6	33.0 ±1.5	1.5 ± 1.3	59	43	54	72
	NM- 103	Rutile (hydrophobic)	Dimethicone 2%	60	21.9 ±1.4	37.9 ±1.6	1.7 ± 1.3	40	33	67	129
	NM- 104	Rutile (hydrophilic)	Glycerine ^e	60	19.0 ±1.5	25.8 ±1.4	1.4±1.3	47	33	60	112
	NM- 105	Rutile-anatase (15-85%)	None ^e	61	20.0 ± 1.3	29.6 ±1.3	1.4±1.2	42	55	90	144



¹ Jan Mast, Keld A. Jensen et al., Nanogenotox Deliverable 4.1, 2013; Tavares et al., 2014

Experimental strategy

Dispersion of NMs according to a standardized protocol





Keld A. Jensen et al., Nanogenotox Deliverable 3, 2011; Tavares et al., 2014

In vitro testing of TiO₂

In vivo testing of TiO₂









No monotonic dose-response relationship; Significant increase in the micronucleus frequency :

*NM-102: 125 μg/ml (*p*=0.038);

***NM-103: 5 e 45 µg/ml** (*p*=0.007 and 0.039)

ΦNM-104: 15 e 45 μg/ml (*p*= 0.037 and 0.048)



1.1. Cytokinesis-block proliferation index in human *lymphocytes*



No Significant decreases of CBPI



1. MN assay in pulmonary cells



Significant increase in the micronucleus frequency in A549 cells exposed to 256 μ g/ml.



Significant (low) increase in the level of DNA breaks in A549 cells exposed

to 128 and 256 µg/ml ; no significant oxidative DNA stress (FPG-modified comet assay)

• NMs obtained under GLP and international benchmarks; good physicochemical characterization; variability associated to experimental conditions minimized:

Differential genotoxicity for closely related NMs observed in human lymphocytes - importance of investigating the toxic potential of each NM individually, instead of assuming a common mechanism and similar genotoxic effects for a set of similar NMs.

Standard genotoxicity tests are useful, and can be applied, for the safety evaluation of nanomaterials – provided that standardized protocols for NM preparation are used, the physicochemical characteristics of NMs are considered.

Predictivity of the *in vitro* genotoxicity assays for *in vivo* situation with NMs?

Remarks



1. Frequency of mutations in the *LacZ* gene recovered from liver and spleen



No mutagenic effects in liver or spleen, 28 days after exposure

Louro et al., EnvironMol Mut (2014)



2. MN in mouse blood immature erythrocytes



3. Comet assay in liver and spleen cells



No induction of DNA damage



In vivo testing of TiO₂

4. Cellular effects in liver cells



Histological analysis of mouse liver



TEM analysis of mouse liver

Persistence of NM in mouse liver and mild inflammatory effects

Louro et al., EnvironMol Mut (2014)

TiO2 NMs – summary of results

NMs	Experimental system	Genotoxicity (MN assay, comet assay or somatic gene mutaions)	Cytotoxicity/ inflammation	
NM-102 NM-103 NM-104 NM-105	Primary lymphocytes	MN assay: Positive, without a dose-response relationship (NM-103, 104) Equivocal (NM-102) Negative (NM-105)	No cell cycle disturbance (CBPI)	
	Alveolar cells type- II (A549)	Equivocal (low +) - comet assay Equivocal – MN assay	Negative	
NM-102	Lung epithelial cell line (BEAS 2B)	Negative	Negative	
	<i>In vivo</i> , Transgenic mice harbouring <i>lacZ</i>	Negative: Comet assay MN assay Somatic mutations in liver	Moderate inflammatory effect in liver NPs accumulation in liver cells	

No systemic mutagenic effects were disclosed for NM-102 in blood, liver and spleen cells of transgenic mice, under the tested conditions

Histological and TEM analyses confirmed the persistence of TiO₂ in liver and showed a moderate inflammatory effect

The integration of the *in vitro* and *in vivo* data strengthens the weight of evidence of an absence of NM-102 primary genotoxicity, although the possibility of a secondary genotoxic effect driven by an inflammatory response within a longer time window or at different doses cannot be excluded. HAZARD ASSESSMENT OF NANOMATERIALS (NMS)











Thank you for your attention!



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